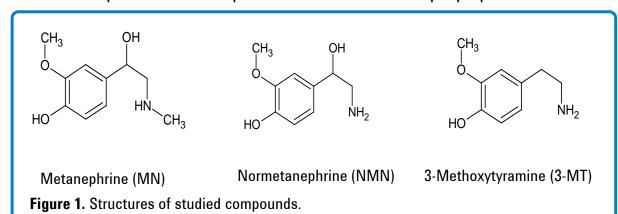
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Introduction

Liquid chromatography triple quadrupole mass spectrometry (LC-MS/MS) is ideally suited for the rapid analysis of multiple analytes. A highly sensitive and specific LC/MS/MS method has been developed for the quantitation of metanephrine, normetanephrine and 3-methoxytyramine in plasma. This method uses a solid phase extraction procedure for efficient sample preparation.



An efficient solid phase extraction (SPE) sample preparation procedure was developed for the simultaneous extraction of metanephrine, normetanephrine and 3-methoxytyramine in plasma. Calibrators were created by spiking clean plasma with various concentrations of each analyte. The chromatographic system consists of a pentafluorophenyl (PFP) column and a mobile phase comprised of methanol and water containing 0.2% formic acid. Quantifier and qualifier MRM transitions were monitored and deuterated internal standards were included for each analyte to ensure accurate and reproducible quantitation.

Experimental

Sample Preparation

Calibrators (Cerilliant, Medical Isotopes) are prepared with clean plasma matrix (Golden West Biologicals). Isotopically labelled Internal standards (Cerilliant, Cambridge Isotopes) and ChromSystems plasma controls were used.

Pretreatment of samples:

0.5 mL plasma, calibrators, QCs Add 50 µL of internal standards mix Add 0.5 mL of 10 mM NH4H2PO4 buffer pH 6.5

- Condition SPE cartridge (SimpliQ WCX, 30 mg, 1 mL) with: 1 mL of MeOH 1 mL of 10 mM NH4H2PO4 buffer pH 6.5
- Add samples
- Wash with 1 mL H2O, 1 mL Methanol, 1 mL 0.2% formic acid in acetonitrile Dry at full vacuum for 5 minutes
- Elute with 2 x 250 μ L of 2% formic acid in acetonitrile. Apply vacuum 5" Hg for 60 seconds Evaporate under nitrogen flow at 40 deg. C Reconstitute with 100 µL of 0.2% formic acid in water Note: use of silanized glassware is recommended for optimum recoveries

LC Method

Agilent 1290 HPLC binary pump, well plate sampler with thermostat, temperaturecontrolled column compartment

Parameter	Value						
Analytical Column	•	Agilent Pursuit 3 PFP, 2x150mm, 3µm PN: A3051150X020					
Guard Column	•	Agilent Meta Guard column Pursuit 3 PFP, 2 mm PN: A3051MG2					
Column Temp	40°C						
Injection Volume	20 µl	20 µl					
Autosampler Temp	4°C	4°C					
Needle Wash	Flush port for 2	Flush port for 20 seconds					
Mobile Phase A	0.2% Formic Ad	0.2% Formic Acid in Water					
Mobile Phase B	0.2 % Formic A	0.2 % Formic Acid in Methanol					
1290 Pump Gradient	Time (min.)	%В	Flow (mL/min.)				
	0.5 1.5 4.0	2 2 60 60 95	0.3 0.3 0.3 0.3 0.3				
	_	95	0.3				
	6.01	95	0.5				
Stop Time	7.5 min.						
Post Time	3 min.						
Table 1. LC Parameters							

Experimental

MS Method

Agilent 6460 QQQ with JetStream technology

lon mode:	AJS ESI(+)
Gas temperature:	350 °C
Drying gas (nitrogen):	5 L/min
Nebulizer gas (nitrogen):	40 psi
Sheath gas (nitrogen):	400 °C
Sheath flow:	11 L/min
Capillary voltage:	3000V
Nozzle voltage:	0V
Q1/Q3 Resolution:	0.7 unit
Dwell time:	40 msec
Delta EMV:	200V

Source conditions and MRM transitions (table 2) were determined and optimized automatically using Agilent Optimizer software.

Compound	Prec Ion	Prod Ion	Frag (V)	CE (V)	CAV (V)
3-Methoxytyramine*	151.1	91.1	135	20	3
3-Methoxytyramine	151.1	119	135	12	3
3-Methoxytyramine-D4	155.1	95.1	135	24	3
Normetanephrine*	166.1	106.1	105	20	3
Normetanephrine	166.1	121	105	20	3
Normetanephrine-D3	169.1	109.1	105	20	3
Metanephrine*	180.1	165.1	120	16	5
Metanephrine	180.1	148.1	120	16	5
Metanephrine-D3	183.1	168.1	120	16	5
Table 2: MRM Transitio					

Results and Discussion

Chromatographic separation of all analytes (figure 2) is achieved in less than four minutes through the use of a pentafluorophenyl (PFP) column. An extra wash can be performed at the end of every run or at the end of a batch for superior ruggedness. Although not measured with this method, the separation of epinephrine and normetanephrine, and the separation of metanephrine and 3methoxytyramine are especially critical since these compounds share common fragments. Without proper separation by retention time, these compounds can cause interferences leading to inaccurate quantitation. The separation also achieved with these compounds is shown (figure 3).

Matrix effect were observed but were compensated for by the internal standards and gave acceptable recovery efficiency (table 3).

Commercially available quality controls (QC) material (ChromSystems) were used to measure the precision of this method. Results (table 4) show excellent precision at both levels.

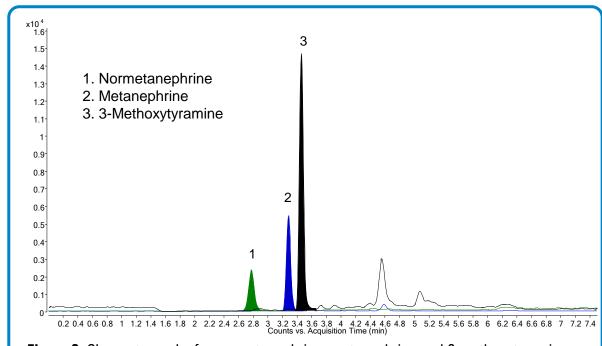
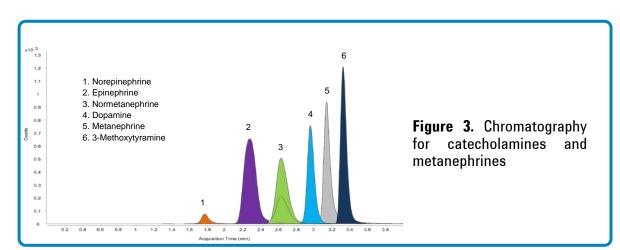


Figure 2. Chromatography for normetanephrine, metanephrine and 3-methoxytyramine



Results and Discussion

Compound	Mati effect (n =	s %	Recovery efficiency % (n = 9)		
	Average	SD	Average	SD	
3-Methoxytyramine	78.3	7.2	99.1	16.9	
Normetanephrine	36.0	13.7	87.7	27.0	
Metanephrine	73.0	9.0	104.4	17.3	

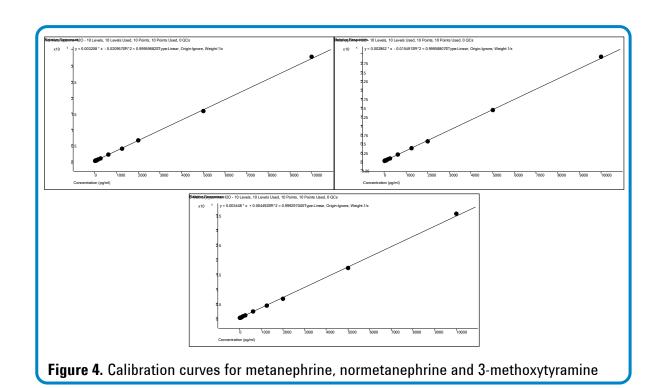
Matrix effect % = B/A *100 Recovery efficiency % = C/B *100

- A: neat standard solutions
- B: plasma extracted then spiked (post-ext)
- C: spiked plasma then extracted (pre-ext)

Table 3: Matrix effects and recovery efficiency

	Level (n=3)		Level 2 (n=3)		
Compound	Measured (pg/mL)	CV (%)	Measured (pg/mL)	CV (%)	
3-Methoxytyramine			1768.9	0.8	
Normetanephrine	95.9	5.1	7369.9	3.0	
Metanephrine	85.0	2.2	1620.8	3.2	

Table 4. Results of ChromSystems controls by LC-MS/MS



Compound	R ²	Concentration (pg/ml)	Concentration (nmol/L)	Accuracy (%) n = 3	Intraday CV(%) n = 3	Interday CV (%) n = 5
	0.9997	15.63	0.09	115.4	2.4	1.9
2 Mothovuturomino		78.13	0.47	97.6	1.8	1.6
3-Methoxytyramine		1250	7.5	100.4	1.1	1.8
		10000	59.8	100.0	0.4	1.0
	0.9996	15.63	0.09	117.3	2.7	4.9
Normatananhrina		78.13	0.43	100.0	2.4	2.5
Normetanephrine		1250	6.8	97.1	2.5	1.8
		10000	54.6	100.4	0.4	0.8
	0.9998	15.63	0.08	116.6	1.7	1.9
Metanephrine		78.13	0.40	96.5	2.5	2.2
inicianchiume		1250	6.3	100.2	1.1	0.9
		10000	50.7	100.3	0.6	0.6

Table 5: Summary of analyte performance.

Conclusions

A robust method for quantifying metanephrine, normetanephrine and 3-methoxytyramine in plasma with excellent reproducibility and accuracy has been developed.

- Whiting, M J. "Simultaneous measurement of urinary metanephrines and catecholamines by liquid
- chromatography with tandem mass spectrometric detection." Ann Clin Biochem 46 (2009): 129-136. Gabler, J. "A sensitive and interference-free liquid chromatography tandem mass spectrometry method for measuring metanephrines in plasma." J Chromatograph Separat Techniq 2012, S2